

Review Article

Heterologous Production of Human Papillomavirus L1 Capsid Protein: Systematic Review and Meta-analysis

Andre Hendrawan & Azzania Fibriani*

School of Life Sciences and Technology, Institut Teknologi Bandung

*) Corresponding author; e-mail: afibriani@sith.itb.ac.id

Received: 2021-11-22

Accepted for publication: 2022-03-22

Abstract

The coverage of HPV vaccination in Indonesia remains low due to the high-cost vaccination. The vaccine prices were affected by the production rate of L1, the active substance of HPV vaccines. L1 has been produced using various organisms with varying L1 production rates and immunogenicity. A systematic review and meta-analysis were conducted to determine the organism producing L1 with the highest production, treatments affecting the L1 expression rate, and immunogenicity (represented by anti-L1 IgG titer in mice). The data of L1 titer, induction period, and IgG titer were extracted from 19 articles that have passed the articles screening. The L1 titer and induction period data were used to calculate the L1 production rate, while the IgG titer was used in the immunogenicity analysis. On a 95% confidence level, the meta-analysis revealed weak evidence that *E. coli* produced L1 at the highest rate. The highest IgG titer was induced using L1 expressed in *Saccharomyces cerevisiae*, albeit insufficient evidence on 95% confidence level. Pearson's correlation analysis showed that the concentration of glucose, IPTG, NH_4^+ , K^+ , Ca^{2+} , Mn^{2+} , Fe^{2+} , Zn^{2+} , $\text{B}_4\text{O}_7^{2-}$, H_2PO_4^- , HPO_4^{2-} , $\text{Mo}_7\text{O}_{24}^{6-}$, and citric acid had a positive correlation with L1 production rate in *E. coli*. The treatment injection doses positively correlated with IgG titer in *S. cerevisiae*. This study reveals the mineral salts as the potential treatments to increase L1 production rates.

Keywords: HPV, L1, L1 production rate, anti-L1 IgG titer, Pearson's correlation analysis

1. Introduction

Cervical cancer remains one of the leading causes of mortality in Indonesia. Global Cancer Observatory: Cancer Today (gco.iarc.fr/today) estimated that in 2020, Indonesia would be the third leading country with the highest cervical cancer incidences — with 36,633 cases with 21,003 mortalities [1].

Human papillomaviruses (HPV), especially the high-risk types, are known as the leading risk factor for cervical cancer [2]. HPV is transmitted through sexual intercourse. HPV infection occurs at a site of epithelial abrasion. Persistent infection by high-risk types induces progression to carcinoma [3].

This day, HPV infections can be prevented through virus-like particle (VLP)-based immunization. However, HPV vaccination coverage in Indonesia remains low. Even though high HPV vaccination coverage has been attained in several cities by 2018, it is not the case for

other regions, especially rural areas [4]. HPV vaccination costs have soared since Indonesia stopped the funding scheme under the cooperation with Global Alliance for Vaccines and Immunizations (GAVI) for the national HPV vaccination in 2019. Thus, HPV vaccines have become more unaffordable for most Indonesian citizens of a lower-middle-income class.

HPV vaccines contain the HPV virus-like protein (VLP) consisting of solely major capsid protein L1 as the active substance [5]. Up until now, the L1 protein has been produced through the expression of the L1 gene using the cells of various organisms [6-10], and the L1 expression rates vary across different kingdoms of life [7, 11]. Moreover, previous experiments on heterologous L1 expression demonstrated that varying nutrient-rich media composition led to varying L1 production rates [12]. Thus, the choice of organism and media formulation are the decisive factors in building the L1 expression system with a higher L1 production rate. The higher the L1

production rate, the lower the selling prices of VLP and, hence, vaccine prices [13]. In addition, the L1 protein produced has to be immunogenic in order to elicit immune responses.

Therefore, this study is aimed to perform a systematic review and meta-analysis to compare the L1 production rate using various organisms and immunogenicity. We also investigated the treatments affecting L1 production rate and immunogenicity.

2. Methodology

2.1. Scope and Search Strategy

The literature search was conducted on September 7, 2021, through PubMed as the search engine. The search terms used were: “human papillomavirus,” AND “L1” AND “expression.” Only the articles in English were included in this systematic review.

2.2. Inclusion and Exclusion Criteria

Firstly, we screened all the articles based on the titles and abstracts and excluded articles unrelated to the heterologous expression of L1. After that, we performed the second stage of screening and included articles containing the data on L1 titer and induction period. We excluded articles that did not include one or two data mentioned above. Articles of co-expressions and chimeric expressions were also excluded, except for the co-expressions of L1 and the chaperone proteins, since those co-expressions increased the L1 titer.

2.3. Meta-analysis

In this study, the immunogenicity is represented by the post-injection anti-L1 IgG titer in mice (*Mus musculus*). The data of L1 titer, induction period, and anti-L1 IgG titer were extracted from 19 articles that have passed the articles screening.

2.4. Calculation of L1 Production Rate

The L1 titer and expression induction period data were extracted from the final selected articles. We calculated the L1 average production rates according to Equation (1).

$$\text{L1 average production rate} = \frac{\text{L1 titre}}{\text{expression induction period}} \quad (1)$$

2.5. Hypothesis Testing

We performed Student's *t*-test using Minitab 17 to determine the statistical significance in comparison of the mean L1 average production rate and the mean anti-L1 IgG titer between each organism.

2.6. Determination of Treatments Affecting L1 Production Rate and Anti-L1 IgG Titer

To seek the treatments affecting the L1 production rate and anti-L1 IgG titer, we determined the Pearson's correlation coefficient for the correlation between each treatment with L1 production rate and anti-L1 IgG titer using Minitab 17.

3. Results and discussion

3.1. Articles Screening

The search yielded 891 papers (1983 to 2021) (Figure 1). The first screening based on titles and abstracts excluded 768 articles, leaving 123 articles. Following the second screening based on the data availability, 104 articles were excluded. Ultimately, the selected 19 articles were used in the meta-analysis.

3.2. Characteristics of Selected Articles

The selected articles were categorized according to the organisms used as the expression hosts (Table 1). The organisms include *Escherichia coli* [6, 14-16], *Saccharomyces cerevisiae* [7, 17-19], *Pichia pastoris* [8, 20], baculovirus-infected *Spodoptera frugiperda* [7, 21-25], *Drosophila* [26], Vero cell [9], MRC-3 cell [9] and tobacco (*Nicotiana benthamiana*) [10].

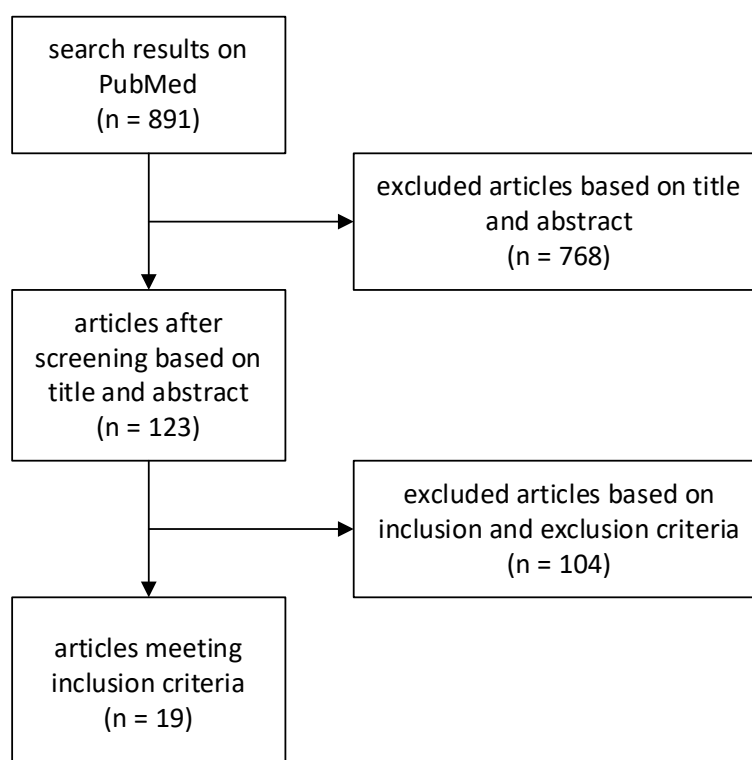
3.3. Production Rate

We extracted 43 data on L1 titer and expression induction period from the selected articles. We calculated each organism's mean L1 average production rate and summarized them in Figure 2. As illustrated in Figure 2, *E. coli* were capable of producing L1 with the highest rate, even though the obtained data were insufficient to support this claim at 95% confidence level due to the high variability of L1 average production rate using *E. coli* (Supplementary Figure 1 and 2). Pearson's correlation analysis for the correlation between 18 treatments and L1 production rate (Supplementary Table 1) revealed that the concentration of glucose, IPTG, NH_4^+ , K^+ , Ca^{2+} , Mn^{2+} , Fe^{2+} , Zn^{2+} , $\text{B}_4\text{O}_7^{2-}$, H_2PO_4^- , HPO_4^{2-} , $\text{Mo}_7\text{O}_{24}^{6-}$ and citric acid had a positive correlation with L1 production rate in *E. coli* ($R = 0.820$ ($P < 0.005$), except for IPTG, to which had a correlation coefficient of $R = 0.619$ ($P < 0.005$)). Those were the components of the R/2 medium used by Bang *et al.* [6]. Bang *et al.* successfully produced 4,6 g/L of L1 capsid using *E. coli* within 4 h expression induction period on this medium, and it was the highest L1 expression rate recorded in this systematic review.

Table 1. Articles classification based on organisms used in experiments.

Organism	Number of Articles
<i>Escherichia coli</i>	4
<i>Saccharomyces cerevisiae</i>	5*
<i>Pichia pastoris</i>	2
baculovirus-infected <i>Spodoptera frugiperda</i>	6*
<i>Drosophila</i>	1
mammalian cells	1
<i>Nicotiana benthamiana</i>	1

* One of the articles compares the L1 expression performance of two organisms, including *S. cerevisiae* and baculovirus-infected *S. frugiperda*.

**Figure 1.** Summary of articles screening.

The L1 production, which attained the highest L1 production rate, is the only study that used the mineral-rich R/2 medium. Bang *et al.* used *E. coli* as the L1 expression host in their study. None of the other studies used similar mineral-rich media as this study. Thus, it is thought that adding the mineral salts into the growth media may escalate the L1 production rates. The mineral salts have been demonstrated to play an essential role in promoting cellular growth. K^+ and Mn^{2+} are the cofactors for several enzymes involved in glycolysis. Ca^{2+} induces the assembly of FtsZ proteins to form protofilaments [27]. The protofilaments can assemble into a Z-ring which drives the membrane invagination during cell division [28]. Fe^{2+} is a ligand for

cytochrome and a precursor for Fe-S proteins. Both of those proteins play an essential role in the electron transport chain in cellular respiration. Zn^{2+} was reported to have a major role as the component of the DNA-binding domain on DNA primase [29]. $B_4O_7^{2-}$ is known as the ligand of autoinducer-2 (AI-2) protein which plays a role in quorum sensing [30] to induce biosynthesis of flagella, chemotaxis, and biofilm formation [31]. $H_2PO_4^-$ and HPO_4^{2-} , aside from being the pH buffer, are needed in the biosynthesis of nucleic acids and phospholipids [32]. $Mo_7O_{24}^{6-}$ is the molybdenum (Mo) source. Mo serves as a ligand for nitrate reductase cofactor, which plays an important role in catalyzing anaerobic nitrate reduction into ammonia [33].

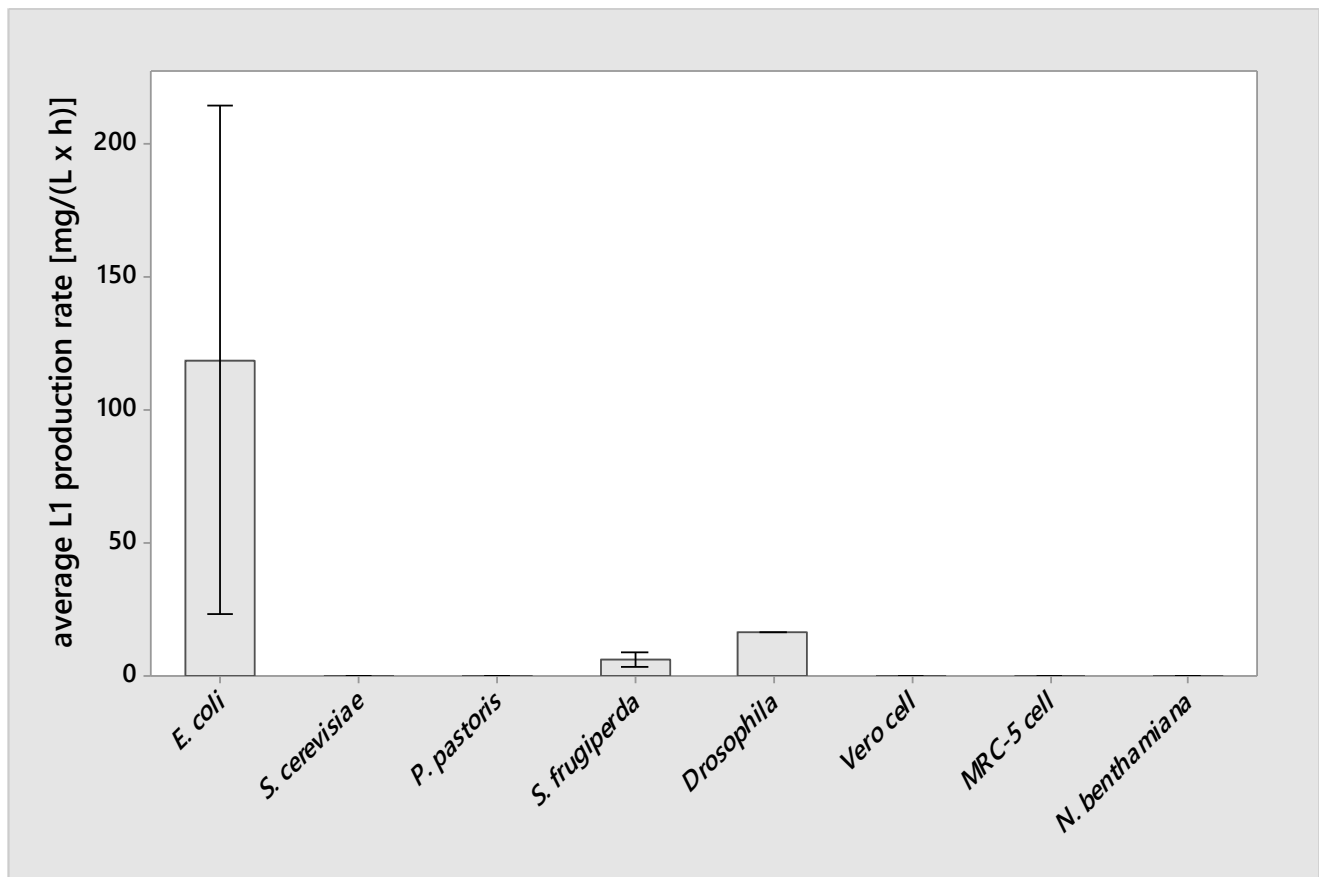


Figure 2. Average L1 production rate for each organism. Bars show the mean of average L1 production rate (not shown for the organism with single datum). Interval bars show the standard deviation. Number of data: *Escherichia coli* ($n = 12$), *Saccharomyces cerevisiae* ($n = 17$), *Pichia pastoris* ($n = 3$), baculovirus-infected *Spodoptera frugiperda* ($n = 6$), *Drosophila* ($n = 1$), Vero cell ($n = 1$), MRC-5 cell ($n = 1$), *Nicotiana benthamiana* ($n = 2$).

Based on this finding, we could suggest the potential idea of implementing the mineral-rich media in L1 production using other organisms. Enriching the media with mineral salts could promote cell viability and thus increase the L1 expression rate using organisms other than *E. coli*. This potential has to be explored further in other organisms.

3.4. Anti-L1 IgG Titer

We extracted 18 data of anti-L1 IgG titer from the selected articles. The mean of anti-L1 IgG titer for each organism was calculated and summarized in Figure 3. As shown in Figure 3, the L1 protein expressed by *Saccharomyces cerevisiae* induced the highest mean of anti-L1 IgG titer in mice, albeit the weak evidence at 95% confidence level [12, 19]. In heterologous L1 expression using *S. cerevisiae*, galactose was added to the media as the inducer [7, 12, 17-19]. However, aside from inducing the heterologous expression, galactose could be utilized as the building block of oligosaccharides in protein glycosylation [34, 35]. In *S. cerevisiae*, the expressed protein is glycosylated [36], and

this might alter the immunogenicity of expressed L1 protein [37]. However, injection doses still became the main factor that affected the immunogenicity, as suggested by Pearson's correlation analysis for 3 treatments (glucose concentration, galactose concentration, and doses) and anti-L1 IgG titer (Supplementary Table 2). The correlation analysis revealed a positive correlation of injection doses with anti-L1 IgG titer ($R = 0.814$ ($P < 0.005$) in *S. cerevisiae*. Although there is evidence that was varying glucose and galactose concentrations led to different anti-L1 IgG titer [12], in this meta-analysis, further observation on scatter diagrams (Supplementary Figures 4 and 5) indicated a weak potential relationship between glucose concentration and anti-L1 IgG titer as well as galactose concentration and anti-L1 IgG titer ($P > 0.05$). Furthermore, the P -value suggested insufficient data available to conclude the relationships. This study calls for more experiments investigating the effect of glucose and galactose on the glycosylation performance and thus anti-L1 IgG titer.

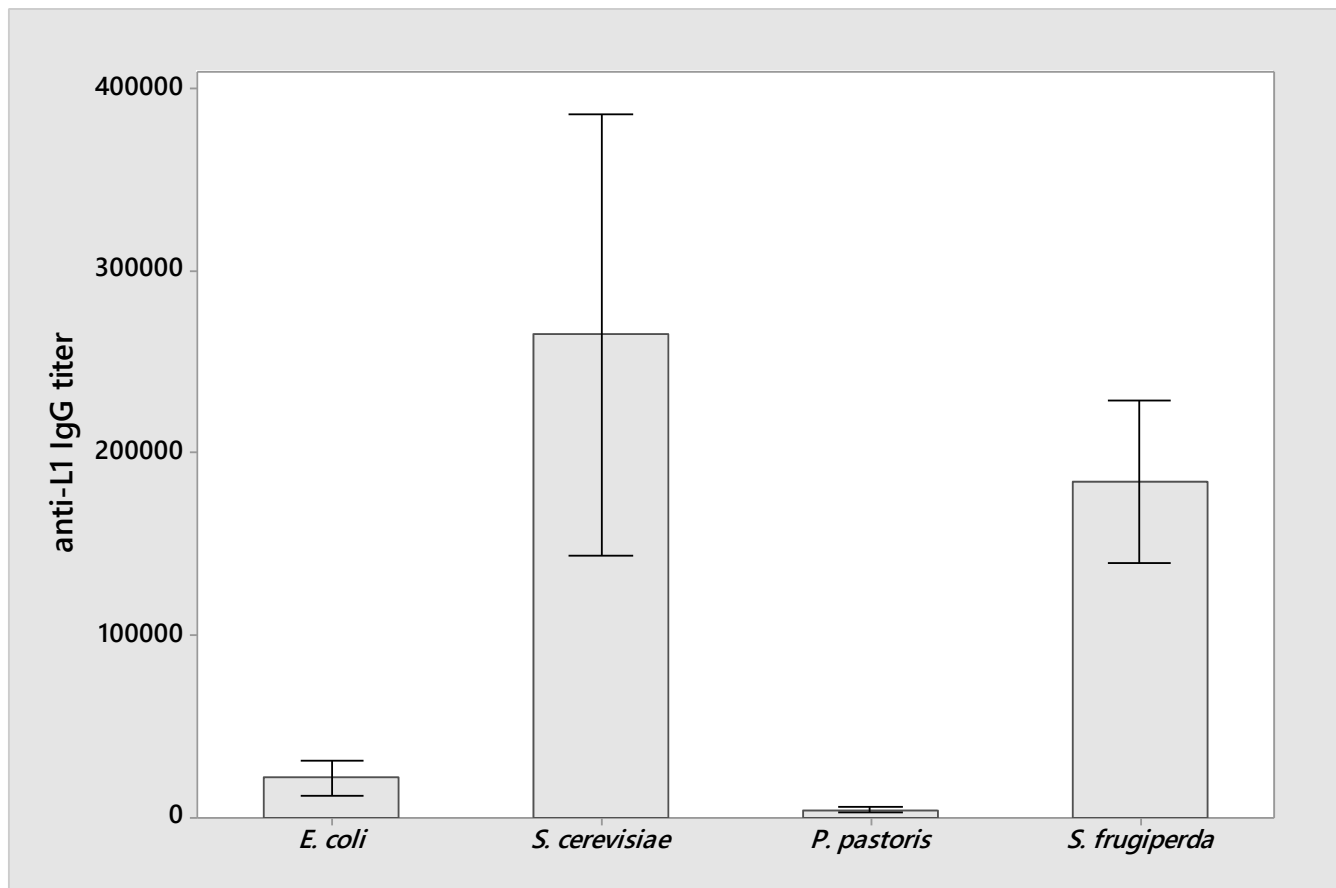


Figure 3. Anti-L1 IgG titer induced by L1 from each organism. Bars show the mean of IgG titer. Interval bars show the standard deviation. Number of data: *Escherichia coli* ($n = 2$), *Saccharomyces cerevisiae* ($n = 10$), *Pichia pastoris* ($n = 2$), baculovirus-infected *Spodoptera frugiperda* ($n = 4$).

4. Conclusion

This is the first study to perform a systematic review and meta-analysis on heterologous L1 expression rate in various organisms and its immunogenicity. The data gathered through this systematic review has yet to provide strong evidence for the statistical significance of different L1 production rates and IgG titers. Meta-analysis revealed weak evidence that *E. coli* produced L1 with the highest rate at 95% confidence level. The highest mean IgG titer was observed on the L1 gene expressed by *Saccharomyces cerevisiae*, albeit the weak evidence at 95% confidence level and its dependence on immunization doses. Pearson's correlation analysis showed that the concentration of glucose, IPTG, NH_4^+ , K^+ , Ca^{2+} , Mn^{2+} , Fe^{2+} , Zn^{2+} , $\text{B}_4\text{O}_7^{2-}$, H_2PO_4^- , HPO_4^{2-} , $\text{Mo}_7\text{O}_{24}^{6-}$, and citric acid had a positive correlation with L1 production rate in *E. coli*, and in *S. cerevisiae*, injection doses had the positive correlation with IgG titer. Additionally, this work provides evidence of the potential role of mineral salts in heterologous L1 expression.

Supplementary

Supplementary material for correlation analysis is provided along with this manuscript.

References

- [1] Ferlay J, Ervik M, Lam F, Colombet M, Mery L, Piñeros M, Znaor A, Soerjomataram I, Bray F. Global Cancer Observatory: Cancer Today. Lyon, France: International Agency for Research on Cancer. Available from: <https://gco.iarc.fr/today> [cited 2021 Dec 1]
- [2] Johansson C, Schwartz S. Regulation of human papillomavirus gene expression by splicing and polyadenylation. *Nature Reviews Microbiology*. 2013 Mar; 11(4): 239-251. DOI: <https://doi.org/10.1038/nrmicro2984>
- [3] Muñoz N, Bosch FX, de Sanjosé S, Herrero R, Castellsagué X, Shah KV, Snijders PJF, Meijer CJLM. Epidemiologic classification of human papillomavirus types associated with cervical cancer. *New England Journal of Medicine*. 2003 Feb;348(6): 518-527. DOI: <https://doi.org/10.1056/NEJMoa021641>
- [4] Ayuningtyas D, Sutrisnawati NND. Indonesia's readiness to implement the HPV vaccine mandatory for school age. *Health*

- Science Journal of Indonesia. 2018 Dec; 9(2): 107-118. DOI: <https://doi.org/10.22435/hsji.v9i2.910>
- [5] Schiller JT, Lowy DR. Understanding and learning from the success of prophylactic human papillomavirus vaccines. *Nature Reviews Microbiology*. 2012 Oct; 10(10): 681-692. DOI: <https://doi.org/10.1038/nrmicro2872>
 - [6] Bang HB, Lee YH, Lee YJ, Jeong, KJ. High-level production of human papillomavirus (HPV) type 16 L1 in *Escherichia coli*. *Journal of Microbiology and Biotechnology*. 2016 Feb;26(2): 356-363. DOI: <https://doi.org/10.4014/jmb.1511.11010>
 - [7] Kim HJ, Cho SY, Park M, Kim H. Comparison of the size distributions and immunogenicity of human papillomavirus type 16 L1 virus-like particles produced in insect and yeast cells. *Archives of Pharmacal Research*. 2018 Apr; 41(5): 544-553. DOI: <https://doi.org/10.1007/s12272-018-1024-4>
 - [8] Rao NH, Babu PB, Rajendra L, Sriraman R, Pang YS, Schiller JT, Srinivasan VA. Expression of codon optimized major capsid protein (L1) of human papillomavirus type 16 and 18 in *Pichia pastoris*; purification and characterization of the virus-like particles. *Vaccine*. 2011 Jul;29(43): 7326-7334. DOI: <https://doi.org/10.1016/j.vaccine.2011.07.071>
 - [9] Zanutto C, Pozzi E, Pacchioni S, Bissa M, Morghen CDG, Radaelli A. Construction and characterisation of a recombinant fowlpox virus that expresses the human papilloma virus L1 protein. *Journal of Translational Medicine*. 2011 Nov;9(190). DOI: <https://doi.org/10.1111/j.1745-7270.2008.00417.x>
 - [10] Regnard GL, Halley-Stott RP, Tanzer FL, Hitzeroth II, Rybicki EP. High level protein expression in plants through the use of a novel autonomously replicating geminivirus shuttle vector. *Plant Biotechnology Journal*. 2009 Nov;8(1): 38-46. DOI: <https://doi.org/10.1111/j.1467-7652.2009.00462.x>
 - [11] Bredell H, Smith JJ, Görgens JF, van Zyl WH. Expression of unique chimeric human papilloma virus type 16 (HPV-16) L1-L2 proteins in *Pichia pastoris* and *Hansenula polymorpha*. *Yeast*. 2018 Apr;35(9): 519-529. DOI: <https://doi.org/10.1002/yea.3318>
 - [12] Kim HJ, Jin Y, Kim HJ. The concentration of carbon source in the medium affects the quality of virus-like particles of human papillomavirus type 16 produced in *Saccharomyces cerevisiae*. *PLoS ONE*. 2014 Apr;9(4): 1-7. DOI: <https://doi.org/10.1371/journal.pone.0094467>
 - [13] Global Alliances for Vaccines and Immunisation. Key concepts: economics of vaccine production. Available from: https://www.who.int/immunization/programmes_systems/financing/analyses/en/briefcase_vacproduction.pdf [Cited 2022 March 9]
 - [14] Pan D, Zha X, Yu X, Wu Y. Enhanced expression of soluble human papillomavirus L1 through co-expression of molecular chaperonin in *Escherichia coli*. *Protein Expression and Purification*. 2015 Dec;120: 92-98. DOI: <https://doi.org/10.1016/j.pep.2015.12.016>
 - [15] Chen Y, Liu Y, Zhang G, Wang A, Dong Z, Qi Y, Wang J, Zhao B, Li N, Jiang M. Human papillomavirus L1 protein expressed in *Escherichia coli* self-assembles into virus-like particles that are highly immunogenic. *Virus Research*. 2016 Apr;220: 97-103. DOI: <https://doi.org/10.1016/j.virusres.2016.04.017>
 - [16] Kelsall SR, Kulski JK. Expression of the major capsid protein of human papillomavirus type 16 in *Escherichia coli*. *Journal of Virological Methods*. 1995 May;53(1): 75-90. DOI: [https://doi.org/10.1016/0166-0934\(95\)00004-e](https://doi.org/10.1016/0166-0934(95)00004-e)
 - [17] Park M, Kim HJ, Kim H. Optimum conditions for production and purification of human papillomavirus type 16 L1 protein from *Saccharomyces cerevisiae*. *Protein Expression and Purification*. 2008 Feb;59(1): 175-181. DOI: <https://doi.org/10.1016/j.pep.2008.01.021>
 - [18] Kim HJ, Kwag H, Jin Y, Kim H. The composition of the carbon source and the time of cell harvest are critical determinants of the final yield of human papillomavirus type 16 L1 protein produced in *Saccharomyces cerevisiae*. *Protein Expression and Purification*. 2011 Jun;80(1): 52-60. DOI: <https://doi.org/10.1016/j.pep.2011.06.010>
 - [19] Kim HJ, Kwag H, Kim H. Codon optimization of the human papillomavirus type 58 L1 gene enhances the expression of soluble L1 protein in *Saccharomyces cerevisiae*. *Biotechnology Letters*. 2012 Nov;35(3): 413-421. DOI: <https://doi.org/10.1007/s10529-012-1097-y>
 - [20] Bazan SB, Chaves AAM, Aires KA, Cianciarullo AM, Garcea RL, Ho PL. Expression and characterization of HPV-16 L1 capsid protein in *Pichia pastoris*. *Archives of Virology*. 2009 Sep;154(10): 1609-1617. DOI: <https://doi.org/10.1007/s00705-009-0484-8>
 - [21] Kirnbauer R, Taub J, Greenstone H, Roden R, Dürst M, Gissman L, Lowy DR, Schiller JT. Efficient self-assembly of human papillomavirus type 16 L1 and L1-L2 into virus-like particles. *Journal of Virology*. 1993 Dec;67(12): 6929-6936. DOI: <https://doi.org/10.1128/JVI.67.12.6929-6936.1993>
 - [22] Wang M, Wang L, Chen L, Han Y, Zou Y, Si J, Song G. Expression of human papillomavirus type 6 L1 and L2 isolated in China and self assembly of virus-like particles by the products. *Acta Biochimica et Biophysica Sinica*. 2003 Jan;35(1): 27-34
 - [23] Zheng J, Ma J, Yang X, Liu H, Cheng H, Si L, Wang Y. Highly efficient and economical baculovirus expression system for preparing human papillomavirus type16 virus-like particle. *Acta Biochimica et Biophysica Sinica*. 2004 Aug;36(8): 548-552. DOI: <https://doi.org/10.1093/abbs/36.8.548>
 - [24] Baek J, Seo J, Kim I, Kim CH. Production and purification of human papillomavirus type 33 L1 virus-like particles from *Spodoptera frugiperda* 9 cells using two-step column chromatography. *Protein Expression and Purification*. 2010 Aug;75(2): 211-217. DOI: <https://doi.org/10.1016/j.pep.2010.08.005>
 - [25] Sun B, Zhao D, Zhang X, Gu T, Yu X, Sun S, Zhao X, Wei L, Liu D, Yan H, Meng X, Kong W, Xu F, Yang P, Jiang C. Development a scalable production process for truncated human papillomavirus type-6 L1 protein using WAVE Bioreactor and hollow fiber membrane. *Applied Microbiology and Biotechnology*. 2015 Oct;100(3): 1231-1240. Available from: DOI: <https://doi.org/10.1007/s00253-015-6974-6>
 - [26] Zheng J, Yang X, Sun Y, Lai B, Wang Y. Stable high-level expression of truncated human papillomavirus type 16 L1 protein in *Drosophila Schneider-2* cells. *Acta Biochimica et Biophysica Sinica*. 2008 May;40(5): 437-442. DOI: <https://doi.org/10.1111/j.1745-7270.2008.00417.x>
 - [27] Yu XC, Margolin W. Ca²⁺-mediated GTP-dependent dynamic assembly of bacterial cell division protein FtsZ into asters and polymer networks in vitro. *EMBO Journal*. 1997 Sep;16(17): 5455-5463. DOI: <https://doi.org/10.1093/emboj/16.17.5455>
 - [28] Margolin W. FtsZ and the division of prokaryotic cells and organelles. *Nature Reviews Molecular Cell Biology*. 2005 Nov;6(11): 862-871. DOI: <https://doi.org/10.1038/nrm1745>

- [29] Kusakabe T, Richardson CC. The role of zinc motif in sequence recognition by DNA primases. *Journal of Biological Chemistry*. 1996 Aug;271(32): 19563-19570. DOI: <https://doi.org/10.1074/jbc.271.32.19563>
- [30] Chen XS, Casini G, Harrison SC, Garcea RL. Papillomavirus capsid protein expression in *Escherichia coli*: purification and assembly of HPV11 and HPV16 L1. *Journal of Molecular Biology*. 2001 Mar;307(1): 173-182. DOI: <https://doi.org/10.1006/jmbi.2000.4464>
- [31] Zohar BA, Kolodkin-Gal I. Quorum sensing in *Escherichia coli*: interkingdom, inter- and intraspecies dialogues, and a suicide-inducing peptide. In: Kalia VC. (eds.) *Quorum Sensing vs Quorum Quenching: A Battle with No End in Sight*. New Delhi, India: Springer India; 2007. p. 85-99
- [32] Madigan MT, Bender KS, Buckley DH, Sattley WM, Stahl DA. *Brock Biology of Microorganisms*. 15th ed., Pearson, 2019
- [33] Lundberg JO, Weitzberg E, Cole JA, Benjamin N. Nitrate, bacteria and human health. *Nature Reviews Microbiology*. 2004 Jul;2(8): 593-602. DOI: <https://doi.org/10.1038/nrmicro929>
- [34] Kornfeld R, Kornfeld S. Assembly of asparagine-linked oligosaccharides. *Annual Review of Biochemistry*. 1985;54: 631-634
- [35] Wildt S, Gerngross TU. The humanization of N-glycosylation pathways in yeast. *Nature Reviews Microbiology*. 2005 Feb;3(2): 119-128
- [36] Conde R, Cueva R, Pablo G, Polaina J, Larriba G. A search for hyperglycosylation signals in yeast glycoproteins. *Journal of Biological Chemistry*. 2004 Oct;279(42): 43789-43798
- [37] Wolfert MA, Boons G. Adaptive immune activation: glycosylation does matter. *Nature Chemical Biology*. 2013 Dec;9: 776-784