

FORMULATION AND CHARACTERIZATION OF NUTMEG (*Myristica fragrans*) ESSENTIAL OIL IN A NANOSTRUCTURED LIPID CARRIER SYSTEM

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ABSTRACT

Essential oil extracted from nutmeg (*Myristica fragrans*) contains various beneficial compounds that can be used in medicines and cosmetics. Nevertheless, nutmeg essential oil (NEO) use in pharmaceuticals is limited by its high lipophilicity and tendency to degrade. Encapsulating NEO into a nanostructured lipid carrier (NLC) can be an alternative to overcome these problems. Thus, this study aimed to develop an NEO formula in an NLC system (NEO-NLC) for topical use, to characterize NEO-NLC, and to elucidate its potential as an antioxidant. The optimization of the NEO-NLC formula included selecting liquid lipid components, adjusting their ratios, choosing a sonicator, selecting a surfactant system, and optimizing sonication amplitude. The optimum NEO-NLC formula was then characterized by determining particle size and distribution, measuring zeta potential, estimating encapsulation efficiency, and evaluating antioxidant activity. The optimum NEO-NLC formula was obtained with a composition of 1.8% cetyl alcohol, 3.0% almond oil, 1.2% NEO, 3.2% Tween 80, 0.8% Plantacare® 1200, and demineralized water. The resulting NEO-NLC exhibited a particle size of 124.4 ± 4.7 nm, a polydispersity index of 0.35 ± 0.05 , a zeta potential of -39.0 ± 3.6 mV, and an encapsulation efficiency of $95.3 \pm 0.6\%$. Encapsulation of NEO into an NLC system resulted in a 2.9-fold increase in antioxidant activity compared to unencapsulated NEO. The developed NEO-NLC showed potential for pharmaceutical or cosmetic raw material use.

Keywords: *Myristica fragrans*, nutmeg essential oil, nanostructured lipid carrier, antioxidant

FORMULASI DAN KARAKTERISASI MINYAK ATSIRI PALA (*Myristica fragrans*) DI DALAM SISTEM NANOSTRUCTURED LIPID CARRIER

ABSTRAK

Minyak atsiri yang diekstraksi dari biji pala (*Myristica fragrans*) mengandung berbagai senyawa bermanfaat yang dapat dimanfaatkan sebagai bahan baku obat maupun kosmetik. Namun, penggunaan minyak atsiri pala (NEO) di bidang farmasi dibatasi oleh sifat lipofilisitasnya yang tinggi dan kecenderungannya untuk terdegradasi. Enkapsulasi NEO ke dalam suatu *nanostructured lipid carrier* (NLC) dapat menjadi alternatif untuk mengatasi permasalahan tersebut. Dengan demikian, penelitian ini bertujuan untuk mengembangkan formula NEO dalam suatu sistem NLC (NEO-NLC) untuk penggunaan secara topikal serta melakukan karakterisasi NEO-NLC termasuk elusidasi potensinya sebagai antioksidan. Optimasi formula NEO-NLC dilakukan melalui pemilihan komponen lipid cair, optimasi rasio komponen lipid, pemilihan tipe sonikator, pemilihan sistem surfaktan, dan optimasi amplitudo sonikasi. Formula NEO-NLC optimum kemudian dikarakterisasi meliputi penentuan ukuran partikel dan distribusinya, pengukuran potensial zeta, dan penentuan efisiensi enkapsulasi, serta elusidasi aktivitas antioksidan. Formula NEO-NLC optimum diperoleh dengan komposisi 1,8% setil alkohol, 3,0% minyak almon, 1,2% MAP, 3,2% Tween 80, 0,8% Plantacare® 1200, dan air demineralisasi. NEO-NLC yang dihasilkan menunjukkan ukuran partikel sebesar 124.4 ± 4.7 nm, indeks polidispersitas 0.35 ± 0.05 , potensial zeta -39.0 ± 3.6 mV, dan efisiensi enkapsulasi $95.3 \pm 0.6\%$. Enkapsulasi NEO ke dalam sistem NLC menghasilkan peningkatan aktivitas antioksidan sebesar 2,9 kali dibandingkan dengan NEO yang tidak dienkapsulasi. NEO-NLC yang dikembangkan menunjukkan potensi untuk digunakan sebagai bahan baku farmasi maupun kosmetik.

Kata Kunci: *Myristica fragrans*, minyak atsiri pala, nanostructured lipid carrier, antioksidan

INTRODUCTION

Nutmeg (*Myristica fragrans*), recognized as one of Indonesia's indigenous plants, presents significant potential as a raw material for pharmaceutical and cosmetic applications. Additionally, essential oil derived from nutmeg (NEO) is particularly notable due to its rich composition of bioactive constituents with proven health benefits. As reported by (Ashokkumar *et al.* 2022), NEO exhibits a diverse range of pharmacological properties, including antibacterial, antioxidant, antifungal, and anti-inflammatory activities. The primary components of NEO that contribute to these beneficial effects include α -pinene, β -pinene, myristicin, 4-terpineol, safrole, sabinene, and limonene (Ansory *et al.* 2020). In addition to its potential applications in the pharmaceutical field, NEO faces several challenges when formulated as a pharmaceutical preparation. These challenges include its high lipophilicity, volatility, and susceptibility to degradation when exposed to environmental factors. Therefore, a delivery system is necessary to protect NEO while enhancing its penetration, particularly for topical applications on the skin.

Nanotechnology has emerged as a groundbreaking field, attracting considerable interest over the last few decades, especially within pharmaceuticals and cosmetics. The concept of nanotechnology was introduced by the Nobel laureate Richard Feynman in 1959, who inspired innovations by discussing the potential of manipulating materials on a small scale (Feynman 1960). Among the fascinating advancements in this field is the nanostructured lipid carrier (NLC), which is an effective vehicle for delivering highly lipophilic substances. NLC represents a significant evolution in lipid nanoparticles, marking it as a second-generation technology that boasts numerous advantages over its predecessor, the solid lipid nanoparticle (SLN).

NLCs are distinguished by their enhanced physical stability, higher drug loading capacity, and minimal drug expulsion during storage, ensuring the therapeutic compounds remain intact and ready for use (Chauhan *et al.* 2020). These advantageous characteristics originate from the innovative design of the NLC, which incorporates liquid lipid components into the carrier system (Muller *et al.*

2011). This unique formulation disrupts the formation of a perfect crystalline structure typically seen in SLNs, resulting in a more versatile and efficient delivery system (Müller, Radtke, and Wissing 2002). Previously, we succeeded in developing an NLC system as the carrier for clove essential oil, which improved clove essential oil's stability and antioxidant activity against fibroblast cells (Satrialdi *et al.* 2024).

In this research, we aimed to enhance the potential application of NEO by encapsulating it within an NLC system. We undertook several optimization processes, which included selecting the appropriate liquid lipid components, adjusting the ratio between solid and liquid lipids, choosing a suitable sonicator, selecting a surfactant system, and optimizing the sonication amplitude. After completing these optimizations, we evaluated the characteristics of the NEO-NLC by assessing particle size and distribution, measuring zeta potential, and estimating encapsulation efficiency. Additionally, we investigated the antioxidant activity of the developed NEO-NLC.

MATERIALS AND METHODS

Materials

The formulation components, including NEO, almond oil, grapeseed oil, virgin coconut oil, sesame oil, were purchased from PT. Darjeeling Sembrani Aroma (West Java, Indonesia), while cetyl alcohol was obtained from Gracefruit Ltd. (Bonnybridge, United Kingdom). The 2,2-diphenyl-1-picrylhydrazyl (DPPH) and ascorbic acid were purchased from PT. Smart-Lab (Banten, Indonesia) and Merck (Rahway, NJ, USA), respectively. All other chemicals and solvents utilized in this research were sourced as commercially available pro-analytical grade.

The fabrication of NEO-NLC

The NEO-NLC was prepared using the emulsification-sonication method, as previously described by (Soleimanian *et al.* 2018), with some minor modifications. First, the lipid components were combined and heated to 60 °C. Simultaneously, the aqueous surfactant system was heated in a separate container until it reached 60 °C. Once both phases reached this temperature, NEO was briefly introduced to the lipid phase,

followed by mixing lipid and water phases using a homogenizer set to 6,600 rpm for 5 minutes. Finally, the mixture was sonicated to produce the NEO-NLC.

Formula optimization of NEO-NLC

Formula optimization was performed to develop the optimal NEO-NLC. This involved screening liquid lipids, optimizing lipid ratios, selecting a sonicator, selecting a surfactant system, and adjusting the sonicator amplitude. Four candidates—almond oil, virgin coconut oil, grapeseed oil, and sesame oil—were tested at a fixed concentration of 1.8%, combined with 3% cetyl alcohol. To stabilize the NEO-NLC, 4% Tween 80 was used, with NEO set at 1.2%. The next step involved optimizing the ratio of cetyl alcohol to the chosen liquid lipid, with variations of 3.0:1.8 and 1.8:3.0. Additionally, we evaluated the type of sonicator used to prepare the NEO-NLC by comparing the particle characteristics obtained from a bath-type sonicator with those from a probe-type sonicator. The impact of adding a co-surfactant was also investigated using two candidates: Plantacare® 1200 and Tego® care 165, combined with Tween 80 in a 1:4 ratio. Additionally, the sonicator amplitude was varied among 40%, 50%, and 60%. The optimal formula was evaluated based on particle size characteristics and its distribution at each stage of the optimization process.

Characterization of NEO-NLC

The optimized NEO-NLC was subjected to particle characterization. This evaluation included determining particle size and distribution, measuring zeta potential, and estimating encapsulation efficiency (EE%). The determination of particle size and distribution was carried out using a particle size analyzer (Beckman Coulter Delsa Nano C, Brea, CA, USA) utilizing the dynamic light scattering method, while the measurement of zeta potential was conducted using the electrophoretic light scattering method on the same instrument. Additionally, to estimate the amount of NEO encapsulated in the NLC system, firstly, the non-encapsulated NEO (free NEO) was separated from the NEO-NLC using centrifugal ultrafiltration at 10,000 rpm for 30 minutes. The concentration of free NEO was then analyzed by

measuring the UV absorption at 266 nm. The encapsulation efficiency was estimated using the following equation (Gomaa *et al.* 2022):

$$EE\% = \frac{\text{initial NEO concentration} - \text{free NEO concentration}}{\text{initial NEO concentration}} \times 100\%$$

Evaluation of antioxidant activity

The antioxidant activity of NEO-NLC was assessed using the DPPH scavenging method, following the technique of (Setiawan *et al.* 2018) with minor modifications. We compared the antioxidant activity of NEO-NLC and free NEO, using ascorbic acid as a control. A 0.2 mM DPPH methanolic solution was mixed with various sample concentrations in a 1:1 volume ratio and incubated in the dark for 30 minutes. After incubation, the absorbance was measured using a UV-Vis spectrophotometer. The antioxidant activity was then calculated using the following equation:

$$Free\ radical\ scavenging\ (\%) = \frac{(DPPH\ absorbance - sample\ absorbance)}{DPPH\ absorbance} \times 100\%$$

The IC₅₀ value, indicating the concentration that scavenges 50% of DPPH, was then calculated from three independent experiments.

Statistical analysis

The quantitative data in this research are presented as average values accompanied by standard deviations (S.D.) calculated from three independent experiments. A T-test was employed to assess the differences between the two groups, while a non-repeated ANOVA followed by the Student-Newman-Keuls (SNK) test was used to differentiate between more than two groups.

RESULT AND DISCUSSION

NEO-NLC was developed using an emulsification process followed by sonication. We began by selecting liquid lipid components to create NEO-NLC, screening four candidates: almond oil, virgin coconut oil, grapeseed oil, and sesame oil. As shown in Table 1, each liquid lipid resulted in different particle characteristics, particularly in particle size and distribution. These differences may arise from variations in the fatty acid composition of the liquid

lipids, differences in their viscosity, and their compatibility with other components in the formulation (Badea *et al.* 2015; Bashiri *et al.* 2020). This optimization process determined that almond oil was the most suitable liquid lipid for preparing NEO-NLC. This conclusion is supported by the resulting particle characteristics, which exhibited a relatively small particle size and a homogeneous

distribution. Additionally, the NEO-NLC produced with almond oil also demonstrated good reproducibility, as indicated by a relatively low standard deviation for particle size and polydispersity index (PDI). Thus, we decided to use almond oil as the liquid lipid component for the following experiments.

Table 1. Particle characteristics of NEO-NLC made from different liquid lipids

| Liquid lipid | Particle size (nm) | PDI |
|--------------------|--------------------|-------------|
| Almond oil | 210.9 ± 16.4 | 0.33 ± 0.03 |
| Virgin coconut oil | 260.3 ± 162.2 | 0.29 ± 0.11 |
| Grapeseed oil | 354.6 ± 67.8 | 0.35 ± 0.17 |
| Sesame oil | 1397.2 ± 1127.8 | 0.63 ± 0.11 |

Note: value = mean ± S.D. (n = 3)

Following the identification of the most appropriate liquid lipid component, we proceeded with our experiment by evaluating the impact of the solid-to-liquid lipid ratio on the characteristics of the particles. We investigated two ratios: 3.0:1.8 and 1.8:3.0. The results summarized in Table 2 demonstrate that the ratio of 1.8:3.0 yielded smaller particle sizes and improved distribution, as indicated by the particle size and PDI values, respectively. A similar trend was observed in our previous research, where an increase in the solid lipid composition resulted in larger NLC particle sizes (Satrialdi *et al.* 2023). This phenomenon can be attributed to the presence of a greater proportion of liquid lipids, which effectively reduces the viscosity and surface tension of the NLC system, thereby facilitating the formation of smaller particle sizes (Hu *et al.* 2005).

In the subsequent optimization process, we examined how different sonication methods affect the characteristics of NEO-NLC particles. We compared two types of sonicators: bath-type and probe-type. A bath-type sonicator generates multiple low-strength cavitation zones, while a probe-type sonicator creates a single, high-strength active zone, concentrating the energy at one point (Jiang *et al.* 2009). According to Figure 1, NEO-NLC produced with a probe-type sonicator had a relatively smaller particle size, improved particle size homogeneity, and greater reproducibility. Based on these findings, we chose to use a probe-type sonicator to prepare NEO-NLC for our next experiments.

Table 2. The effects of lipid composition on characteristics of NEO-NLC

| Cetyl alcohol (%) | Almond oil (%) | Particle size (nm) | PDI |
|-------------------|----------------|--------------------|-------------|
| 3.0 | 1.8 | 210.9 ± 16.4 | 0.33 ± 0.03 |
| 1.8 | 3.0 | 170.1 ± 35.6 | 0.31 ± 0.01 |

Note: value = mean ± S.D. (n = 3)

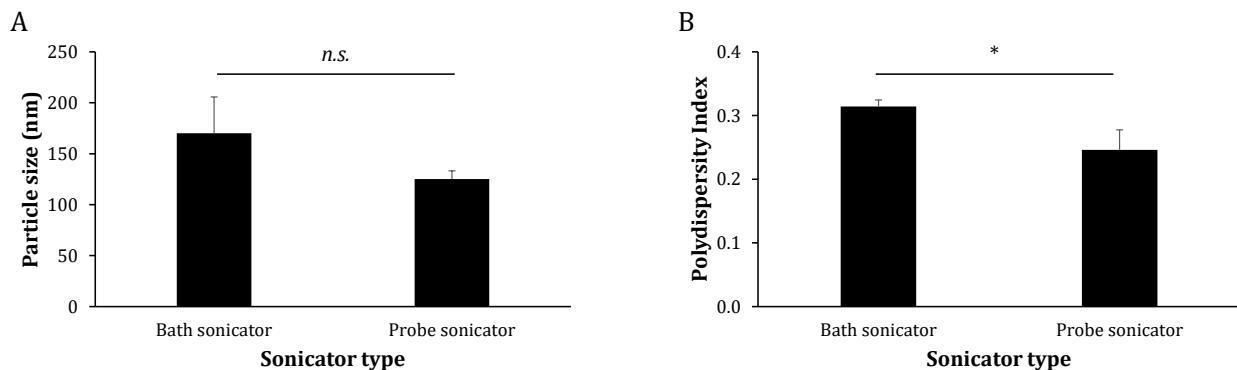


Figure 1. The effects of sonication type on (A) the particle size and (B) its distribution of the NEO-NLC. Value represented as the mean with S.D. from 3 independent experiments (n.s. = not significant; * $p<0.05$ by two-tail unpaired T-test).

After completing the above optimization processes, we proceeded with the NEO-NLC formulation by examining the changes in particle size after storing it for one day at a temperature of 4 °C. We observed a significant increase in the particle size of the NEO-NLC after this storage period (Table 3). This increase suggests that the resulting NEO-NLCs exhibit low stability. One possible explanation for this issue is that the single surfactant system cannot adequately stabilize the NEO-NLC. To address this, we investigated the effect of adding a co-surfactant on the stability of the NEO-NLC. We

combined Tween 80 with Plantacare® 1200 or Tego® Care 165 in a ratio of 4:1. As presented in Table 3, the inclusion of both Plantacare® 1200 and Tego® Care 165 enhanced the stability of the NEO-NLC, as shown by the minimal changes in particle size and distribution. However, we noted that some lipids adhered to the vial wall in aggregate form when NEO-NLC was prepared using Tego® Care 165 as a co-surfactant. Consequently, we used Plantacare® 1200 as the co-surfactant for preparing the NEO-NLC.

Table 3. The screening of surfactant system and its effects on the stability of the NEO-NLC

| Surfactant system | Particle size (nm) | | PDI | |
|---------------------------|--------------------|----------------|----------------|----------------|
| | H ₀ | H ₁ | H ₀ | H ₁ |
| Tween 80 | 125.2 ± 8.0 | 343.9 ± 305.0 | 0.25 ± 0.03 | 0.27 ± 0.06 |
| Tween 80-Plantacare® 1200 | 96.3 ± 4.3 | 95.0 ± 12.9 | 0.34 ± 0.02 | 0.35 ± 0.02 |
| Tween 80-Tego® Care 165 | 112.0 ± 9.9 | 120.8 ± 17.2 | 0.34 ± 0.04 | 0.29 ± 0.05 |

Note: value = mean ± S.D. (n = 3)

We then examined how sonication amplitude affects the properties of NEO-NLC. We tested three levels of sonication: 40%, 50%, and 60%. Figure 2 shows that increasing the sonication amplitude decreases the particle size. This happens because a higher amplitude generates more energy, which breaks up the particles. However, too much energy can create bubbles and lead to aggregation. Thus, sonication amplitude is crucial when making an NLC system. Based on the results in Figure 2, we chose a sonication amplitude of 50% to prepare NEO-NLC. The best formula for producing NEO-NLC is presented in Table 4, which uses a probe sonicator set at 50% amplitude.

The optimized formulation of NEO-NLC underwent a comprehensive characterization process. As presented in Table 4, the NEO-NLC demonstrated a particle size of approximately 125 nm, accompanied by an acceptable PDI value, indicating a homogeneous particle distribution. Furthermore, the zeta potential, which reflects the surface charge of the particles, revealed a notably high negative charge of approximately -39.0 mV. More than 95% of NEO was successfully encapsulated within the NLC system. Collectively, the characteristics of the resulting NEO-NLC suggest its potential efficacy as a topical agent. We then evaluated the antioxidant activity of NEO-NLC compared to non-encapsulated NEO using the DPPH scavenging

method. DPPH is a free radical that can react with an antioxidant agent, forming a reduced form of DPPH (Munteanu and Apetrei 2021). Our findings indicated that encapsulating NEO in an NLC system improved its antioxidant activity by 2.9-fold, as reflected in the IC_{50} value (Table 5). The observed increase in antioxidant activity can be attributed to the inclusion of almond oil in the NEO-NLC formula. Almond oil is renowned for its rich antioxidant content, which neutralizes free radicals and

reduces oxidative stress (Csakvari *et al.* 2019). This oil has a synergistic effect with NEO, which enhances the overall antioxidant scavenging efficacy of the formula. Although the IC_{50} of NEO-NLC is relatively high compared to ascorbic acid, the developed NEO-NLC shows promise as a therapeutic strategy for various skin-related disorders, including antibacterial, antifungal, and anti-inflammatory applications.

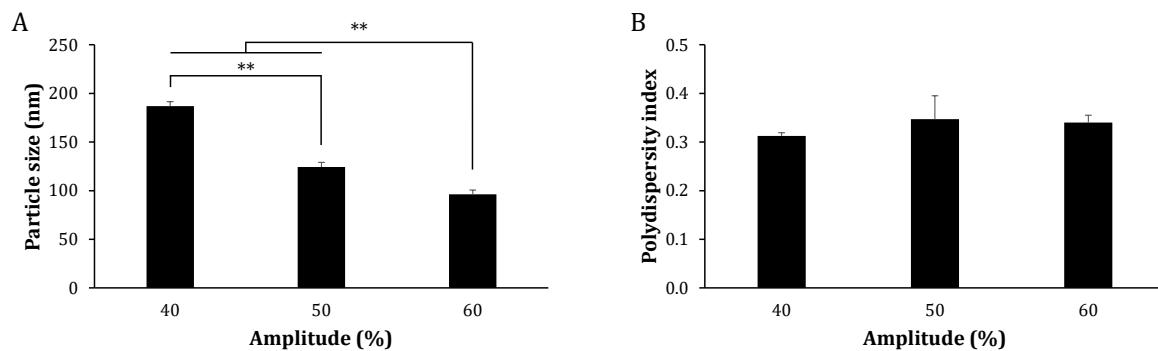


Figure 2. The effects of sonication amplitude on (A) the particle size and (B) its distribution of the NEO-NLC. The error bars indicate S.D. from three independent experiments (not significant for polydispersity index and ** $p<0.01$ by non-repeated ANOVA followed by SNK Test).

Table 4. The optimized formulation of NEO-NLC and its characteristics

| Material | Concentration (%) | Particle characteristics |
|---------------------|-------------------|--|
| NEO | 1.2 | Particle size: 124.4 ± 4.7 nm |
| Cetyl alcohol | 1.8 | PDI: 0.35 ± 0.05 |
| Almond oil | 3.0 | ζ -potential: -39.0 ± 3.6 mV |
| Tween 80 | 3.2 | EE: $95.3 \pm 0.6\%$ |
| Plantacare® 1200 | 0.8 | |
| Demineralized water | Up to 100.0 | |

Note: value = mean \pm S.D. ($n = 3$)

Table 5. DPPH radical scavenging activity of NEO and NEO-NLC

| Sample name | IC_{50} (μ g/mL) |
|---------------|-------------------------|
| Ascorbic acid | 10.9 ± 0.7 |
| NEO | 6770.8 ± 260.5 |
| NEO-NLC | 2337.3 ± 101.7 |

Note: value = mean \pm S.D. ($n = 3$)

CONCLUSIONS

In summary, we have developed an NLC system for encapsulating NEO with the following composition: 1.8% cetyl alcohol, 3.0% almond oil, 1.2% NEO, 3.2% Tween 80, 0.8% Plantacare® 1200, and demineralized water. The resulting NEO-NLC

displayed a particle size of 124.4 ± 4.7 nm, a polydispersity index of 0.35 ± 0.05 , a zeta potential of -39.0 ± 3.6 mV, and an encapsulation efficiency of $95.3 \pm 0.6\%$. We observed that the NEO-NLC exhibited a 2.9-fold increase in antioxidant activity compared to free NEO. Furthermore, the NEO-NLC showed promising potential as a therapeutic

strategy for various skin-related disorders and in cosmetic applications.

REFERENCES

- Ansory HM, Sari EN, Nilawati A, Handayani S, Aznam N, 2020, Sunscreen and Antioxidant Potential of Myristicin in Nutmeg Essential Oils (*Myristica fragrans*), The proceedings of the 2nd Bakti Tunas Husada-Health Science International Conference (BTH-HSIC 2019), 2019, Tasikmalaya.
- Ashokkumar K, Simal-Gandara J, Murugan M, Dhanya MK, Pandian A, 2022, Nutmeg (*Myristica Fragrans* Houtt.) Essential Oil: A Review on Its Composition, Biological, and Pharmacological Activities, *Phytother Res* 36(7):2839–51, doi: 10.1002/ptr.7491.
- Badea G, Lăcătușu I, Badea N, Ott C, Meghea A, 2015, Use of Various Vegetable Oils in Designing Photoprotective Nanostructured Formulations for UV Protection and Antioxidant Activity, *Ind Crops Prod* 67:18–24, doi: 10.1016/j.indcrop.2014.12.049.
- Bashiri S, Ghanbarzadeh B, Ayaseh A, Dehghannya J, Ehsani A, Ozyurt H, 2020, Essential Oil-Loaded Nanostructured Lipid Carriers: The Effects of Liquid Lipid Type on the Physicochemical Properties in Beverage Models, *Food Biosci* 35:100526, doi: 10.1016/j.fbio.2020.100526.
- Chauhan I, Yasir M, Verma M, Singh AP, 2020, Nanostructured Lipid Carriers: A Groundbreaking Approach for Transdermal Drug Delivery, *Adv Pharm Bull* 10(2):150–65, doi: 10.34172/apb.2020.021.
- Csakvari AC, Lupitu A, Bungău S, Gîtea MA, Gîtea D, Țiț DM, Copolovici L, Nemeth S, Copolovici D, 2019, Fatty Acids Profile and Antioxidant Activity of Almond Oils Obtained From Six Romanian Varieties, *Farmacia* 67(5):882–87, doi: 10.31925/farmacia.2019.5.19.
- Feynman RP, 1960, There's Plenty of Room at the Bottom, *Engineering and Science* 23(5):22–36.
- Gomaa E, Fathi HA, Eissa NG, Elsabahy M, 2022, Methods for Preparation of Nanostructured Lipid Carriers, *Methods* 199: 3–8, doi: 10.1016/j.ymeth.2021.05.003.
- Muller RH, Shegokar R, Keck CM, 2011, 20 Years of Lipid Nanoparticles (SLN & NLC): Present State of Development & Industrial Applications, *Curr Drug Discov Technol* 8(3): 207–27, doi: 10.2174/157016311796799062.
- Hu, Fu-Qiang, Sai-Ping Jiang, Yong-Zhong Du, Hong Yuan, Yi-Qing Ye, and Su Zeng, 2005, Preparation and Characterization of Stearic Acid Nanostructured Lipid Carriers by Solvent Diffusion Method in an Aqueous System, *Colloids Surf B Biointerfaces* 45(3–4):167–73, doi: 10.1016/j.colsurfb.2005.08.005.
- Jiang J, Oberdörster G, Biswas P, 2009, Characterization of Size, Surface Charge, and Agglomeration State of Nanoparticle Dispersions for Toxicological Studies, *J Nanopart Res* 11(1):77–89, doi: 10.1007/s11051-008-9446-4.
- Müller RH, Radtke M, Wissing SA, 2002, Solid Lipid Nanoparticles (SLN) and Nanostructured Lipid Carriers (NLC) in Cosmetic and Dermatological Preparations, *Adv Drug Deliv Rev* 54(SUPPL.):131–55, doi: 10.1016/S0169-409X(02)00118-7.
- Munteanu IG, Apetrei C, 2021, Analytical Methods Used in Determining Antioxidant Activity: A Review, *Int J Mol Sci* 22(7), doi: 10.3390/ijms22073380.
- Satrialdi, Putri PA, Lumintang YA, 2023, PENGEMBANGAN FORMULA NANOSTRUCTURED LIPID CARRIER (NLC) SEBAGAI PEMBAWA MINYAK ATSIRI MELATI (*Jasminum Officinale L.*) SERTA POTENSI AKTIVITAS ANTIOKSIDAN, *Majalah Farmasi dan Farmakologi* 27(2), doi: 10.20956/mff.v27i2.27395.

15. Satrialdi, Husna NH, Rihad RQ, Utami RA, 2024, The Incorporation of Clove Essential Oil into Nanostructured Lipid Carrier for Improvement of the Delivery and Antioxidant Effects on the Fibroblast Cells, *Pharm Nanotechnol* 12, doi: 10.2174/0122117385304491240320040711.
16. Setiawan F, Yunita O, Kurniawan A, 2018, Uji Aktivitas Antioksidan Ekstrak Etanol Kayu Secang (*Caesalpinia Sappan*) Menggunakan Metode DPPH, ABTS, Dan FRAP, *Media Pharmaceutica Indonesiana* 2(2): 82-89.
17. Soleimanian Y, Goli SAH, Varshosaz J, Sahafi SM, 2018, Formulation and Characterization of Novel Nanostructured Lipid Carriers Made from Beeswax, Propolis Wax and Pomegranate Seed Oil, *Food Chem* 244: 83-92, doi: 10.1016/j.foodchem.2017.10.010.